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Citation for final published version:

Zahra, Rabaab, Javeed, Saba, Malala, Bibi, Babenko, Dmitriy and Toleman, Mark A ORCID: <https://orcid.org/0000-0002-9497-0512> 2018. Analysis of Escherichia coli STs and resistance mechanisms in sewage from Islamabad, Pakistan indicates a difference in E. coli carriage types between South Asia and Europe. Journal of Antimicrobial Chemotherapy 73 (7) , pp. 1781-1785. 10.1093/jac/dky109 file

Publishers page: <http://dx.doi.org/10.1093/jac/dky109>
<<http://dx.doi.org/10.1093/jac/dky109>>

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Analysis of *Escherichia coli* sequence types and resistance mechanisms in sewage from Islamabad, Pakistan indicates difference in *E. coli* carriage types between S. Asia and Europe

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Running title: Carriage *E. coli* sequence types in Islamabad, Pakistan.

1 **Abstract:**

2 **Objectives:** To discover the *E. coli* carriage sequence types and associated
3 resistance mechanisms in the community in Islamabad, Pakistan by analysis of *E.*
4 *coli* isolates in sewage.

5 **Methods:** 110 *E. coli* were isolated from sewage across the city of Islamabad
6 without antibiotic bias and confirmed as *E. coli* by MALDI-TOF. Isolates were
7 characterized by CH typing and cgMLST. Resistance mechanisms, virulence
8 genes, phylotypes and plasmid incompatibility types were determined in a
9 subset of isolates by in-silico analysis. The genomic position of *bla*_{CTX-M-15} was
10 determined using S1 PFGE, probing and Nanopore minion sequencing.

11 **Results and conclusions:** The most prevalent sequence types were ST394, ST10
12 and ST648 accounting for 39% of all isolates collected and were found at many
13 sites across Islamabad. Carbapenemase genes were absent and only a single
14 isolate of ST131 was found. *qnrS1* and *bla*_{CTX-M-15} were the most prevalent
15 resistance mechanisms with *bla*_{CTX-M-15} penetrating many sequence types and
16 31% of all collected isolates. However, the majority of the successful sequence
17 types were *bla*_{CTX-M-15} negative indicating that resistance is not the main driver of
18 prevalence. 23% of *bla*_{CTX-M-15} genes were chromosomally encoded and large
19 *ISEcP1* mediated insertions included *qnrS1* and several plasmid genes. In all
20 chromosomally encoded isolates no plasmid copies of *bla*_{CTX-M-15} were found. The
21 most prevalent ST (ST394) contained many Enteroaggregative *E. coli* (EAEC)
22 virulence genes and the *fimH30* variant allele previously linked to the success of
23 ST131.

24

25

26 **Introduction:**

27 Antibiotic resistance in *Escherichia coli* is a major concern as *E. coli* is both a
28 leading cause of human infection and an ever-present gut colonist. Carriage of
29 ESBL *E. coli* can lead to difficult to treat urinary tract infections (UTIs), delay
30 appropriate therapy and lead to poor outcomes.¹

31 The *E. coli* species consists of many different sequence types (>7,000
32 <http://pubmlst.org/databases.shtml>) and strains.² Individual strains vary in
33 ability to cause disease and to resist antibiotics. Several molecular
34 methodologies have enabled comparisons of strains between diverse geographic
35 locations including: MLST; *fimH/fumC* (CH typing); ³ Phylo-typing ⁴ and WGS.
36 Comparisons have revealed that resistance is associated with a small number of
37 successful strains and sequence types, for example ST131 *E. coli* belonging to
38 phylogroup B2 ⁵ and *E. coli* ST101 belonging to the phylogroup B1.⁶ We have also
39 seen expansion of ST131 ESBL *E. coli* causing severe infections.⁷ A recent study
40 investigating ESBL *E. coli* in the UK found ST131 was the primary cause of
41 cephalosporin resistant bacteraemia's and the most prevalent ST in faeces and
42 sewage (H. Cadden, P. Cleary, M. Day, M. Doumith, M. Ellington, N. Elviss, J.
43 Findlay, K. Hopkins, B. Jones, D. Livermore, L. Randall, C. Teale, M. Toleman, D.
44 Wareham, C. Wiuff, N. Woodford unpublished results). This suggests that
45 analysis of *E. coli* in sewage is a good proxy for gut carriage in the community.

46 In this study, we sought to determine the carriage *E. coli* sequence
47 types/resistance mechanisms by analysis of *E. coli* collected without antibiotic
48 selection from 18 sewage outfall sites across the city of Islamabad.

49

50

51 **Materials and Methods:**

52 **Collection of samples:** Samples (30 mL) were collected from 18 sites in 9
53 sectors of Islamabad (Figure S1).

54 **Bacterial isolation and identification:** Samples were centrifuged, pelleted and
55 re-suspended in 0.5mL LB broth. 50 µL was spread on MacConkey plates without
56 antibiotic and grown at 37°C overnight. Ten colonies with typical *E. coli*
57 morphology were randomly collected from each site with a total of 110 colonies
58 confirmed as *E. coli* by MALDI-TOF.

59 **Detection of *bla*_{CTX-M} and *bla*_{NDM-1} genes:** PCR utilized custom primers and
60 ReddyMix Extensor PCR Master Mix 1 (Thermo Scientific) with appropriate
61 controls.

62 **Two Locus CH typing:** *fimH* and *fumC* genes were amplified by PCR as described
63 by Weissman.³ *fumC* and *fimH* alleles were assigned using
64 (<http://mlst.warwick.ac.uk/mlst/dbs/Ecoli>) and (<https://cge.cbs.dtu.dk/services/FimTyper/>)
65 websites, respectively.

66 ***bla*_{CTX-M-15} genomic location:** S1 PFGE was performed as described.⁸ The probe
67 was amplified using primers CTXMF/R (GTTACGCTGATGGCGACGGC,
68 ACGCTAATACATCGCGACGGC) and radio-labeled using P³² dCTP as described
69 previously.⁸

70 **Genomic DNA extraction:** Genomic DNA was extracted using the Qiagen
71 genomic DNA kit.

72 **Miseq Sequencing:** DNA libraries were prepared using the NexteraXT sample
73 kit and sequenced (20-30× coverage) with a standard 2x100 base protocol on a
74 MiSeq Instrument (Illumina, San Diego, CA, USA) in house at Cardiff.

Sequence type and resistance/virulence gene detection: MLST was determined with StringMLST using short read data in fastq format and Ridom SeqSphere+ (version 3.5.0) using assembled data in fasta format. *E. coli* strains were clustered based on cgMLST typing with SeqSphere+. A core genome MLST (cgMLST) scheme was based on *E. coli* ATCC25922 with 1907 targets. Antimicrobial resistance genes were detected using CLC-Biogenomic workbench. Plasmid and virulence genes were detected using Plasmid finder (<https://cge.cbs.dtu.dk/services/PlasmidFinder/>); and Virulence finder (<https://cge.cbs.dtu.dk/services/VirulenceFinder/>).

Minion sequencing: Isolates with chromosomally encoded *bla*_{CTX-M-15} were sequenced using the nanopore minion RAD-002 rapid sequencing kit following isolation of high molecular weight DNA by the CTAB (hexadecyltrimethylammonium bromide) method.⁹ Single reads were used as scaffold to assemble miseq data and annotated using geneious.

Phylo-group analysis: The *E. coli* phylogroups were determined with in-silico searches for *chuA*, *yjaA*, *tspE4.C2*, *arpAgpE*, *tnpAgpC* using geneious software based on the Clermont method.⁴

99 **Results and discussion**

100 This study was designed to discover *E. coli* carriage ST and resistance
101 mechanisms in Islamabad. We collected *E. coli* at sewage outfalls across the city
102 and isolated 110 *E. coli* strains without antibiotic selection. PCR indicated 34
103 isolates carried *bla*_{CTX-M-15} and a single isolate carried *bla*_{CTX-M-27}. All isolates were
104 *bla*_{NDM-1} negative and carbapenem susceptible indicating low carbapenemase
105 carriage rates in Islamabad as compared to other S. Asian sites.^{10 11 12 13}
106 CH clonotyping revealed 24 known MLST in the collection of 110 *E. coli*'s. These
107 were *fumC*-*fimH*: 35-30 (ST394); 11-54 (ST10); 4-0 (ST648); 65-32/ 54-0
108 (ST1431) and 7-54 (ST45) which accounted for 15%, 13%, 7%, 3.6% and 2.7%
109 of isolates respectively. Two isolates of clonotypes, *fumC*-*fimH*: 4-27 (ST88); 29-
110 38 (ST156); 6-0 (ST688); 7-54 (ST2325); 636-34 (ST5176) and 11-24 (ST43)
111 which equates to 1% prevalence each and single isolates of each clonotype
112 including: 11-27 (ST93); 40-30 (ST131); 6-31 (ST154); 4-32 (ST155); 23-54
113 (ST205); 11-69 (ST216); 4-61 (ST224); 7-25 (ST398); 136-331 (ST543); 65-32
114 (ST1128) and 27-0 (ST4121). Details of location, numbers of ST and respective
115 clonotypes are shown in Figure S1. Of 110 *E. coli* isolates, 33 were whole genome
116 sequenced (WGS) including examples of all MLST sequence types and multiple
117 examples of common ones (Table S1). The in silico MLST agreed 100% with
118 MLST derived from CH clonotyping. Most frequently found sequence types were
119 ST394, ST10, ST648 and ST1431 found at 44%, 44%, 28% and 11% of the
120 sampling sites, respectively (Figure S1). Interestingly, ST131 was particularly
121 rare (a single isolate <1%). Isolation of *E. coli* without antibiotic selection
122 enabled us to measure penetration of resistance mechanisms through this
123 species. *bla*_{CTX-M-15} is highly prevalent worldwide¹⁴ and colonized 23% of *E. coli*

isolates in Islamabad including 63% of MLST. However, it was found only in 14%, 24% and 50% of individual isolates of the most prevalent types: ST648; ST394 and ST10, respectively. This indicates that *bla*_{CTX-M-15} *E. coli* are overall less fit than parental isolates and that the prevalence of individual *E. coli* ST is not directly related to cephalosporin resistance.

WGS revealed all resistance and virulence mechanisms in 33 isolates including 24 positive and 9 negative CTX-M isolates (Table S1). The *qnrS1* gene was most common, found in 80 isolates by PCR. This was often associated with *bla*_{CTX-M-15} (91%). Other prevalent mechanisms were: *dhfr* genes found in 60%, *bla*_{TEM-1B} in 33%, *tetA* in 42%, *sul2* in 33%, *sul1* 21% and *tetB* 9% of WGS isolates. Other mechanisms were rarely found: *bla*_{OXA-1} in 3; *bla*_{TEM-199} in 2; and *bla*_{TEM-158}, *bla*_{CMY-42}, and *bla*_{CMY-44} and in individual isolates Table S1.

WGS detected 19 different virulence genes (Table S1). Overall the majority of isolates were commensals with few virulence factors belonging to phylogroups A, C and B1. The ST394 isolates belonged to phylogroup E and harboured entero-aggregative virulence factors typical of the EAEC group of *E. coli*. ST394 is associated with diarrheal disease but also commonly recovered from healthy people and has been implicated in acute and persistent sporadic diarrhea, and outbreaks, in both industrialized and developing countries.¹⁵ ST131 and ST648 were the only EXPEC pathogens recovered belonging to phylogroups B2 and D, respectively. They are known to cause UTI's and sepsis, though they do not carry the classical EXPEC virulence gene repertoire.¹⁶

S1 PFGE analysis revealed 1-4 plasmids present in each strain. Plasmids were of a range of incompatibility groups (Table S1). The *bla*_{CTX-M-15} plasmids ranged in size from 50-150 kb. We found *bla*_{CTX-M-15} genes on the chromosome in 8 isolates

(23%) and 6 ST. We determined the insertion sites in 4 isolates including identical sites in ST394 and a unique insertion in ST4121 (Figure S2). In all cases movement to the chromosome was *ISEcP1* mediated and confirmed by the presence of direct repeats generated at the site of insertion (Figure S2). Notably these movement events not only transposed *bla*_{CTX-M-15} to the chromosome but also sections of a common plasmid containing *qnrS1*. Movement to the chromosome is typically associated with subsequent loss of the *bla*_{CTX-M-15} plasmid as evidenced by lack of *bla*_{CTX-M-15} harboring plasmids in these strains. This process likely enhances the fitness of the strain overall.¹⁷

We used cgMLST to relate the various *E. coli* to each other (Figure 1). ST were correctly grouped together. The Clermont PCR phylogroup analysis gave good agreement in general with cgMLST. Interestingly, cgMLST aligned the ST648 isolates as group F instead of D, confirming the recent observation of Johnson¹⁸ and highlighting the superior discriminatory power of cgMLST.

Analysis of the *fimH* data revealed that several strains were *fimH* null. This included all ST648 isolates and is typical of this ST.⁶ Its prevalence as an EXPEC pathogen suggests that another adhesion is substituting for *fimH*. The worldwide success of the *bla*_{CTX-M-15} gene in ST131 *E. coli* has been closely linked to a highly adherent *fimH* variant allele, *fimH30*.⁵ In this study despite ST131 being particularly rare, *fimH30* was the second most prevalent variant of *fimH* and was present in all ST394 isolates. The possession of *fimH30* by ST394 is likely advantageous. However, the fact that ST131 *fimH30 E. coli* is particularly rare suggests that other as yet unknown factors are responsible for the success of ST394 in Islamabad.

Acknowledgements: This study was presented as a poster at the British Society of Antimicrobial Chemotherapy Spring meeting in March 2017 “Association of Antibiotic genes with Successful ST of *Escherichia coli* in The Environment.”

Funding: This project was funded by a Commonwealth Academic Staff fellowship grant (PKCF-2016-156) awarded to Dr Rabaab Zahra.

Transparency Declaration: All authors have no conflicting interests to declare.

Figure and Table legends:

Figure 1. cgMLST of Islamabad sewage isolates.

cgMLST was generated using a core genome of 1907 target genes identified in each sequenced isolate using Ridom SeqSphere+ software. Phylogroup analysis was based on in silico PCR using targets identified by Clermont *et al* 2013.

Figure 2. Chromosomal insertion sites of *bla*_{CTX-M-15}.

Chromosomal insertion sites were identified using a combination of miseq and Nanopore Minion single reads sequences. (A) represents the insertion site found in the ST4121 isolate 8 and (B) represents the insertion site found in ST394 isolates 2, 12 and 15. The 11,389bp insertion found in isolates 2, 12 and 15 included *bla*_{CTX-M-15} and *qnrS1* and was identical to the first 11,389bp of the 23,174bp insertion in ST4121 isolate 8. The larger insertion in ST4121 isolate 8 included additional *tetA* and *tetR* resistance genes as well as several other genes of plasmid origin. The chromosomal insertion in the ST394 isolates was in a low GC % intergenic region between *ydcS* and *ydcR* genes encoding a putative DNA binding transcriptional regulator and a putative spermidine putrescine transporter subunit, respectively. The insertion in the ST4121 isolate was in a hydrolase gene. Each insertion had the mobile element *ISEcp1* at the LH terminus and was flanked by target site direct repeats of TATGA for isolate 8 and TTAAA for isolates 2, 12 and 15. This figure appears in colour in the online version of *JAC* and in black and white in the printed version of *JAC*.

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S2 legend

(Figure 3). This insertion was 11,398 bp and was *ISEcP1* mediated with a typical 5bp target site duplication of host DNA adjacent to the *ISEcP1* left hand terminus (TATGA) and a surrogate *ISEcP1* right hand terminus at the insertion site boundaries. The insertion included *bla*_{CTX-M-15} and *qnrS1* resistance genes as well as several partial or complete transposon genes and was similar to *bla*_{CTX-M-15} containing sections of several plasmids suggesting a plasmid origin. The chromosomal insertion in ST4121 included an identical section of DNA adjacent to *ISEcP1* but was considerably longer including 23,174 bp and included *tetA* and *tetR* genes in addition to *bla*_{CTX-M-15} and *qnrS1* genes as well as several other genes of plasmid origin. This insertion was also *ISEcP1* mediated with a typical direct repeat of target DNA at the insertion site (TTAAAA), which was at position 112bp of a hydrolase gene (Figure 3).